

# Angiogenesis in health and disease

Peter Carmeliet

**Blood vessels constitute the first organ in the embryo and form the largest network in our body but, sadly, are also often deadly. When dysregulated, the formation of new blood vessels contributes to numerous malignant, ischemic, inflammatory, infectious and immune disorders. Molecular insights into these processes are being generated at a rapidly increasing pace, offering new therapeutic opportunities that are currently being evaluated.**

## Vessel growth: modes and impact on health

Small blood vessels consist only of endothelial cells (ECs), whereas larger vessels are surrounded by mural cells (pericytes in medium-sized and smooth muscle cells (SMCs) in large vessels). Vessels can grow in several ways. Vasculogenesis refers to the formation of blood vessels by endothelial progenitors, angiogenesis and arteriogenesis refer to the sprouting and subsequent stabilization of these sprouts by mural cells, and collateral growth denotes the expansive growth of pre-existing vessels, forming collateral bridges between arterial networks. Both capillary angiogenesis and arterial growth are targets for therapy, as distal capillaries distribute the flow while proximal arterioles provide bulk flow to the tissue. When vessel growth is dysregulated, it has a major impact on our health and contributes to the pathogenesis of many disorders, some quite unexpected. Indeed, a long list of disorders is characterized or caused by excessive angiogenesis. Historically, the best known are cancer, psoriasis, arthritis and blindness, but many additional common disorders such as obesity, asthma, atherosclerosis and infectious disease are included, and the list is still growing (Table 1). Several congenital or inherited diseases are also caused by abnormal vascular remodeling (Table 1). In addition, insufficient vessel growth and abnormal vessel regression not only cause heart and brain ischemia, but can also lead to neurodegeneration, hypertension, pre-eclampsia, respiratory distress, osteoporosis and other disorders (Table 2). Few other processes have as daunting an impact as angiogenesis on the well-being of so many people worldwide. Recent advances in the understanding of molecular, genetic and cellular mechanisms of vessel growth and their possible implications for medicine will be discussed in this overview.

## Endothelial progenitors

For many years, the prevailing dogma stated that vessels in the embryo developed from endothelial progenitors, whereas sprouting of vessels in the adult resulted only from division of differentiated ECs. Recent evidence, however, indicates that endothelial progenitors contribute to vessel growth both in the embryo and in ischemic, malignant or inflamed tissues in the adult, and can even be therapeutically used to stimulate ves-

sel growth in ischemic tissues, a process termed 'therapeutic vasculogenesis'<sup>1-3</sup> (Fig. 1). ECs differentiate from angioblasts in the embryo<sup>4</sup> and from endothelial progenitor cells (EPCs), mesoangioblasts, multipotent adult progenitor cells, or side-population cells in the adult bone marrow<sup>1,5</sup>. EPCs can also contribute to vessel growth by releasing angiogenic growth factors<sup>6</sup>. ECs may also share a common origin with blood cells in the embryo and arise from the hemangioblast<sup>4</sup>. Endothelial and hematopoietic progenitors and their descendents share common markers, are affected by common signals, and influence each other. For instance, hematopoietic stem cells (HSCs) bud from hemogenic ECs in the embryo, and HSCs and leukocytes stimulate angiogenesis partly by releasing angiogenic factors or transdifferentiating to ECs<sup>7-10</sup>. Identification of the signals that recruit or differentiate these progenitors offers opportunities to manipulate their contributions to vascular growth. Vascular endothelial growth factor (VEGF), placental growth factor (PlGF, a homolog of VEGF), angiopoietin (Ang)-1, inhibitor of differentiation (Id) proteins, cytokines, and other signals have a role<sup>9-12</sup>. Overall, the functional contribution of EPCs and HSCs to pathological angiogenesis still remains largely undefined (see accompanying review in this issue<sup>13</sup>).

## Vascular cell specification

Endothelial progenitors differentiate to mature ECs, but not all ECs are alike. One well-known anatomical and physiological distinction between vessels is that of arteries and veins. Not only do they differ in the blood pressure they sustain and the thickness of their SMC coat, but their ECs and SMCs also have a distinct identity and origin. For instance, SMCs surrounding some thoracic vessels are derived from neural crest, whereas coronary SMCs are derived from epicardium, and other SMCs arise from mesenchyme<sup>14</sup>. Little is known about the various pathways specifying the identity of arterial and venous SMCs, but recent genetic studies offer insight into the signals controlling arterial and venous identities of ECs. The Notch pathway, with its ligands (Delta-like-4, Jagged-1 and Jagged-2) and receptors (Notch-1, Notch-3 and Notch-4), promotes arterial fate of ECs by repressing venous differentiation<sup>15,16</sup>. Sonic Hedgehog and VEGF act upstream, whereas Gridlock probably acts downstream of Notch to determine arterial fate, even before the onset of flow<sup>16,17</sup>. ECs can differentiate into either arterial or venous ECs in embryonic development, in the neonatal retina and even in the adult heart, indicating that ECs have a remarkable phenotypic plasticity<sup>18,19</sup>. Selective use of arterial or venous ECs or their precursors may offer opportunities for therapeutic

Center for Transgene Technology and Gene Therapy, Flanders Interuniversity Institute for Biotechnology, KULeuven, Campus Gasthuisberg, Herestraat 49, B-3000, Leuven, Belgium.

E-mail: peter.carmeliet@med.kuleuven.ac.be





tial vasculogenesis. Notch signaling, however, is also critical for proper maintenance of arteries. Mutations of the SMC-specific Notch-3 receptor, which disrupt SMC anchorage to the extracellular matrix (ECM) and impair SMC survival, cause degeneration of cerebral arterioles, leading to cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy<sup>20</sup>. Besides Notch, bone-marrow tyrosine kinase and neuropilin-1 (a VEGF<sub>164</sub>-specific receptor) also influence arterial specification<sup>18</sup>. By secreting VEGF, peripheral nerves codetermine arterial differentiation, providing a molecular explanation as to why arteries and nerves often run alongside each other in the body<sup>21</sup>.

Blood vessels in various tissues have specialized functions, and ECs are therefore equipped with distinct properties—there might be even as many different EC types as there are organs in the body (see accompanying review in this issue<sup>22</sup>). What determines this EC heterogeneity and organ-specific angiogenesis? First, the expression and activity of general angiogenic factors such as VEGF or Ang-1 varies greatly in different tissues. Low-permeability tumors overexpress Ang-1 or underexpress VEGF (or both), whereas high-permeability tumors lack Ang-1 (ref. 23). Another example is the effect of Ang-1, which stimulates angiogenesis in the skin but suppresses vascular growth in the heart<sup>19,24</sup>. Second, organ-specific angiogenic factors determine the angiogenic switch, but in a restricted manner in particular organs (for example, blood vessel/epicardial substance and fibulin-2 in the heart, and endocrine gland VEGF and prokineticin-2 in endocrine glands<sup>25</sup>). Such organ-specific molecules hold great promise for use in developing safer angiogenic therapies. Tumor vessels also change their phenotype and express new addresses ('vascular zip codes'), which are absent or barely detectable in quiescent vessels<sup>26</sup>. Some vessels are not even lined by ECs: cytotrophoblasts line the maternal spiral arteries during normal placentation (a process termed 'pseudovasculogenesis'), SMCs line the neointima when re-endothelialization after vessel injury is incomplete, and malignant cells line some tumor vessels (a process called 'vascular mimicry'<sup>27</sup>).

**Vascular boundaries and polarity**

After endothelial progenitors differentiate into ECs and form a primitive vascular labyrinth, further remodeling of such primitive vessels into a more complex network requires the demarcation of arterial and venous boundaries, as well as the establishment of vascular polarity (Fig. 1). The Eph-Ephrin system is involved in the organization of such vascular boundaries. EphrinB2 marks arterial ECs and SMCs, whereas EphB4, a receptor for EphrinB2, marks only veins. EphrinB2-EphB4 signaling is critical for the establishment of arterial and venous identities, and participates in the formation of arteriovenous anastomoses by arresting EC migration at the arterial-venous interface<sup>28–30</sup>. Capillaries were long considered to lack any identity, but EphrinB2 expression extends into capillaries midway between terminal arterioles and post-capillary venules, indicating that they are either arterial or venous. As development proceeds, EphrinB2 expression extends also to SMCs in arteries. In pathological angiogenesis, ECs of some new vessels also express EphrinB2, contrary to the dogma that tumor vessels arise exclusively from postcapillary venules<sup>31,32</sup>.

Very little is known about vascular polarity, yet many vessels, such as the large thoracic vessels, develop in an asymmetric pattern and are only present in the left or right side of the body.

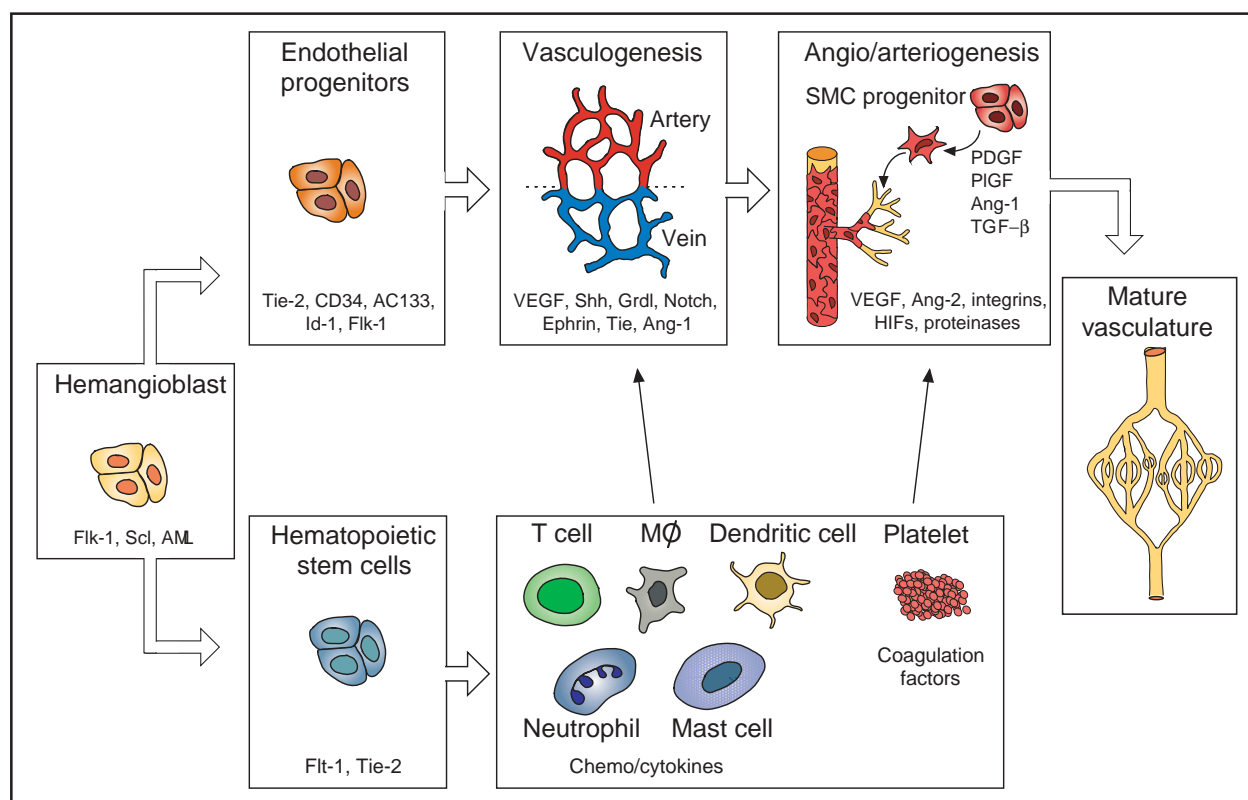
The embryonic pharyngeal arch arteries (PAA) initially develop symmetrically, but are subsequently remodeled asymmetrically into various large thoracic arteries. Because of its complexity, this process is often derailed, giving rise to congenital vascular malformations. Hotspots of VEGF expression around the PAAs are essential for their asymmetric remodeling. When VEGF expression is dysregulated, the left-side fourth PAA abnormally regresses, whereas the right-side fourth PAA, predetermined to regress otherwise, persists as a right-side aorta, giving rise to the typical vascular malformations and birth defects found in DiGeorge syndrome<sup>33</sup>. A combinatorial role of Ang-1 and the Tie-1 receptor seems to be essential in establishing the right-side venous system<sup>34</sup>. There are many vascular malformations, especially in neural tissue, that may result from 'misguiding' and aberrant patterning, but their etiology remains largely enigmatic. Another intriguing question is whether homeobox genes determine vascular identity, boundaries, polarity and patterning.

**Angiogenesis and arteriogenesis**

The nascent vascular bed expands by sprouting and matures into a system of stable vessels (Fig. 1). Hypoxia is an important stimulus for expansion of the vascular bed. Initially, cells are oxygenated by simple diffusion of oxygen, but when tissues grow beyond the limit of oxygen diffusion, hypoxia triggers vessel growth by signaling through hypoxia-inducible transcription factors (HIFs; see accompanying review in this issue<sup>35</sup>). HIFs upregulate many angiogenic genes, but the induction of VEGF is perhaps the most remarkable—up to 30-fold within minutes. VEGF stimulates physiological and pathological angiogenesis in a strict dose-dependent manner and is therefore currently being evaluated for pro- and antiangiogenic therapy (see accompanying review in this issue<sup>36</sup>). Loss of a single allele causes embryonic vascular defects<sup>37,38</sup>, and reduction of VEGF levels by only 25% impairs spinal cord perfusion and results in motor neuron degeneration, reminiscent of amyotrophic lateral sclerosis<sup>39</sup>. PlGF, which binds Flt-1, enhances angiogenesis but only under pathological conditions. It amplifies VEGF-driven angiogenesis in part through a unique cross-talk between Flt-1 and Flk-1 (refs. 12,40). The role of VEGFB in angiogenesis remains to be determined. Besides VEGF family members, numerous other molecules have been documented to regulate EC growth, including growth factors, chemokines, cytokines, lipid mediators, hormones and neuropeptides (see below).

**Table 1 Diseases characterized or caused by abnormal or excessive angiogenesis**

Organ	Diseases in mice or humans
Numerous organs	Cancer (activation of oncogenes; loss of tumor suppressors); infectious diseases (pathogens express angiogenic genes <sup>112</sup> , induce angiogenic programs <sup>113</sup> or transform ECs <sup>114</sup> ); autoimmune disorders (activation of mast cells and other leukocytes)
Blood vessels	Vascular malformations (Tie-2 mutation <sup>68</sup> ); DiGeorge syndrome (low VEGF and neuropilin-1 expression <sup>33</sup> ); HHT (mutations of endoglin or ALK-1 (ref. 69)); cavernous hemangioma (loss of Cx37 and Cx40 (ref. 44)); atherosclerosis; transplant arteriopathy
Adipose tissue	Obesity (angiogenesis induced by fatty diet; weight loss by angiogenesis inhibitors <sup>115</sup> )
Skin	Psoriasis, warts, allergic dermatitis, scar keloids, pyogenic granulomas, blistering disease, Kaposi sarcoma in AIDS patients <sup>114</sup>
Eye	Persistent hyperplastic vitreous syndrome (loss of Ang-2 (refs. 65,116) or VEGF164 (ref. 18)); diabetic retinopathy; retinopathy of prematurity; choroidal neovascularization (TIMP-3 mutation <sup>51</sup> )
Lung	Primary pulmonary hypertension (germline BMPR-2 mutation; somatic EC mutations <sup>73,75,76</sup> ); asthma; nasal polyps
Intestines	Inflammatory bowel and periodontal disease, ascites, peritoneal adhesions
Reproductive system	Endometriosis, uterine bleeding, ovarian cysts, ovarian hyperstimulation <sup>25</sup>
Bone, joints	Arthritis, synovitis, osteomyelitis, osteophyte formation <sup>12</sup>



**Figure 1** Formation of a vascular network. Endothelial progenitors differentiate to arterial and venous ECs, which assemble in a primitive capillary plexus. Vessels then sprout and become stabilized by SMCs, differentiating from their progenitors. HSCs contribute to angiogenesis directly and indirectly, by differentiating to leukocytes or platelets. A partial list of molecules is indicated; see text for additional information. Shh, Sonic hedgehog; Grd1, Gridlock; M $\phi$ , macrophage; AML, acute myeloid leukemia; Scl, stem cell leukemia.

ECs are elongated, thin and fragile cells, yet they build channels that do not collapse and that efficiently distribute blood to the various parts of the body. They also have long half-lives of several years, but when triggered are capable of rapidly sending out sprouts in a coordinated and directional manner. How can they possess all these qualities? It is partly because cells within the vessel wall communicate with each other and with cells inside and outside the vessel lumen. They sense changes in blood flow and pressure, and dynamically interact with the internal cytoskeleton and surrounding ECM, all in an integrated manner. Vascular cells are equipped with a set of molecules that allow them to perform these functions (see below). In quiescent vessels, vascular endothelial cadherin in adherens junctions and claudins, as well as occludin and JAM-1 in tight junctions, provide mechanical strength and tightness and establish a permeability barrier. These molecules do not only serve as 'mechanical zippers', but also transmit crucial signals for endothelial survival and other functions<sup>41</sup>. When ECs migrate during vessel sprouting, these contacts are transiently dissolved but later re-established, once ECs assemble a new sprout. Interrupting this cycle disrupts vessel assembly in tumors<sup>42</sup>. VEGF loosens, whereas Ang-1 tightens these contacts; the therapeutic potential of the latter is currently being evaluated in conditions of sepsis, inflammation, injury, stroke and cancer<sup>43</sup>. Homotypic ECs contacts through CD31 (PECAM) and intercellular communication through connexins (Cx) in gap junctions are also crucial for vessel formation and maintenance, as the loss of both Cx37 and Cx40 causes cavernous hemangiomas, and deficiency in Cx43 dysregulates coronary artery formation<sup>44</sup>.

The ECM provides necessary contacts between ECs and the surrounding tissue, and thus prevents vessels from collapsing. In quiescent vessels,

a basement membrane of collagen IV, laminin and other components encases vascular cells; pericytes and ECs are even embedded in the same basement membrane. An interstitial matrix of collagen I and elastin between vascular cells further provides visco-elasticity and strength to the vessel wall. The ECM also regulates the formation of new vessel sprouts. When vascular cells migrate to form new sprouts, this matrix network is not only proteolytically broken down, but its composition is also altered. Proteinases expose new cryptic epitopes in ECM proteins (such as in collagen IV) or change their structure (fibrillar versus monomer collagen), which induce EC and SMC migration<sup>45</sup>. In addition, a provisional matrix of fibronectin, fibrin and other components provides a support scaffold, guiding ECs to their targets. Integrins are cell-surface receptors of specific ECM molecules that, by bidirectionally transmitting information between the outside and inside of vascular cells, assist vascular cells to build new vessels in coordination with their surroundings<sup>46,47</sup>. The  $\alpha_v\beta_3$  and  $\alpha_v\beta_5$  integrins have long been considered to positively regulate the angiogenic switch, because their pharmacological antagonists suppress pathological angiogenesis. Genetic deletion studies suggest, however, that vascular integrins inhibit angiogenesis by suppressing VEGF- and Flk-1-mediated EC survival, by *trans*-dominantly blocking other integrins or by mediating the antiangiogenic activity of thrombospondins (TSPs) and other angiogenesis inhibitors (such as tumstatin, endostatin, angiostatin and PEX). It remains to be determined whether and under what conditions integrins have positive or negative roles in angiogenesis.

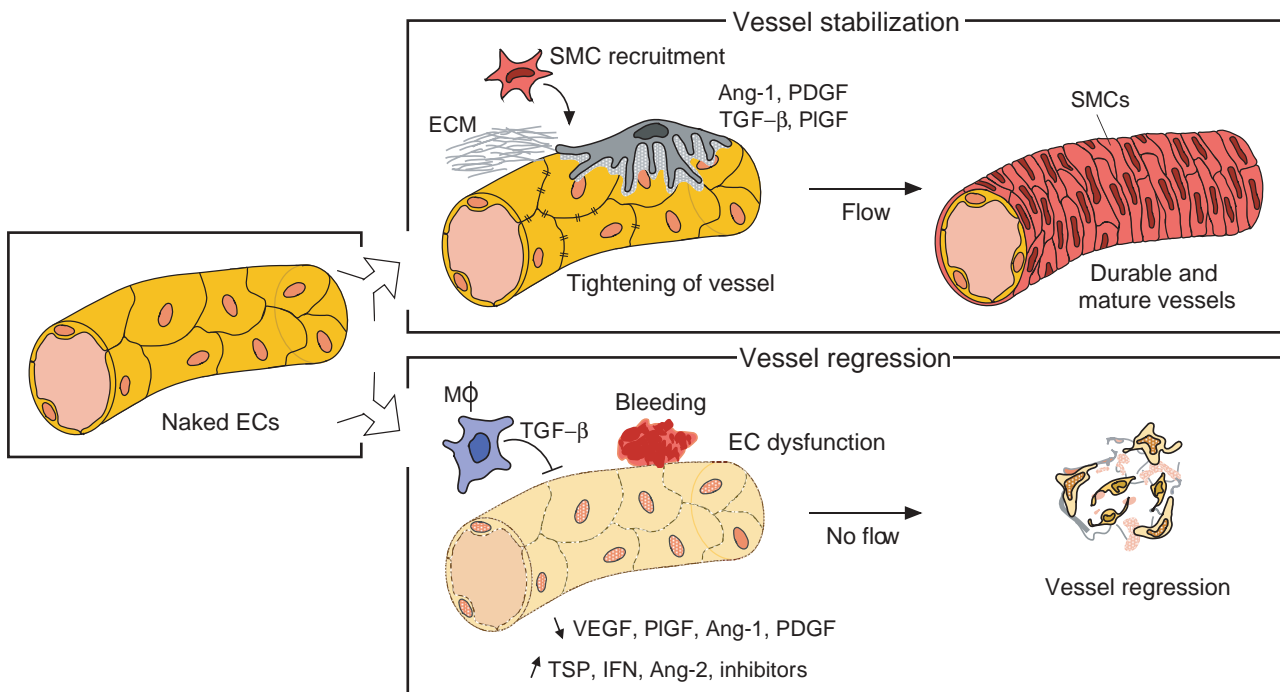
Remodeling of the ECM during vessel sprouting requires breakdown by proteinases, including plasminogen activators (such as urokinase plasminogen activator (uPA) and its inhibitor, PAI-1), matrix metallopro-

teinases (MMPs and tissue inhibitors of metalloproteinases (TIMPs)), heparinases, chymases, tryptases and cathepsins<sup>48–50</sup>. Proteinases also facilitate EC sprouting by liberating matrix-bound angiogenic activators (basic fibroblast growth factor (FGF), VEGF and transforming growth factor (TGF)- $\beta$ ) and proteolytically activating angiogenic chemokines (such as IL-1 $\beta$ ). Their activity is, however, not always related to proteolysis, as shown for uPA receptor and TIMP-3 (refs. 51,52). When considering the critical role of the ECM in vessel growth and maintenance, it is conceivable that proteolytic remodeling of the ECM must occur in a balanced manner. Insufficient breakdown prevents vascular cells from leaving their original position, but excessive breakdown removes critical support and guidance cues for migrating ECs and, in fact, inhibits angiogenesis<sup>50,53</sup>. Proteinases can also have a role in the resolution of angiogenesis, as they liberate matrix-bound inhibitors (TSP-1, canstatin, tumstatin, endostatin and platelet factor (PF)-4) and inactivate angiogenic cytokines (such as stromal cell-derived factor-1). These pleiotropic activities may explain why proteinases and their receptors and inhibitors often have activities that are context- and concentration-dependent. It may also explain why an inhibitor such as PAI-1 is a predictor of poor, not good, clinical outcome for many cancers<sup>50,53</sup>.

Establishment of a functional vascular network further requires that nascent vessels mature into durable vessels (Fig. 2). The association of pericytes and SMCs with newly formed vessels regulates EC proliferation, survival, migration, differentiation, vascular branching, blood flow and vascular permeability (see accompanying review in this issue<sup>54</sup>). Platelet-derived growth factor (PDGF)-BB and its receptor, PDGFR- $\beta$ , have essential roles in the stabilization of nascent blood vessels by recruiting PDGFR- $\beta$ -positive mesenchymal progenitors. Dropout or insufficient recruitment of mural cells results in EC growth, permeability, fragility, vessel enlargement, bleeding, impaired perfusion and hypoxia in

embryos lacking PDGF-B<sup>55</sup>, in retinas of diabetics, in tumors<sup>56</sup> and in hemangiomas, which are the nonmalignant vascular tumors that rapidly enlarge in infants and often spontaneously regress<sup>57</sup>. The subsequent increase in VEGF further aggravates vascular permeability and edema, and promotes hemangioma formation. In contrast, a combination of PDGF-BB and VEGF results in the formation of more mature vessels than monotherapy with either factor, a finding relevant for future development of therapeutic angiogenesis strategies<sup>58</sup>. PDGF-CC and PDGF-DD also promote angiogenesis, but their roles remain less well characterized<sup>59</sup>.

Another signaling system involved in vessel maintenance, growth and stabilization is the Tie-2 receptor, which binds the angiopoietins (Ang-1 and Ang-2). Unlike Ang-2, which activates Tie-2 on some cells but blocks Tie-2 on others, Ang-1 consistently activates Tie-2. Even though trapping angiopoietins suppresses pathological vascularization<sup>60</sup>, their role is pleiotropic and context-dependent. Ang-1 stimulates vessel growth in skin, ischemic limbs, gastric ulcers and in some tumors<sup>23,61</sup>, presumably because it is an EC survival factor and mobilizes EPCs and HSCs<sup>62</sup>. But Ang-1 also suppresses angiogenesis in tumors and the heart<sup>19,63</sup>. Although it is still not entirely understood, the antiangiogenic effect of Ang-1 may relate to the fact that vessels must loosen up before ECs can migrate; if vessels are too tight, vessel sprouting may be impeded. Ang-1 tightens vessels by affecting junctional molecules<sup>43</sup> and by promoting the interaction between ECs and mural cells as an adhesive protein and recruiting pericytes<sup>64</sup>. Ang-2 has been proposed to stimulate the growth of immature (SMC-poor) tumor vessels by loosening endothelial-periendoneothelial cell interactions and degrading the extracellular matrix, thereby antagonizing Ang-1 (refs. 63,65). The angiogenic activity of Ang-2 seems to be contextual as well, however. Ang-2 synergizes with VEGF to stimulate angiogenesis in the heart<sup>19</sup> but, when insufficient angiogenic



**Figure 2** Vessel maintenance versus vessel regression. Nascent vessels initially only consist of ECs. Upper panel: vessel maturation requires a mix of angio- and arteriogenic factors for a sufficient duration, so that ECs can tighten up and become covered by mural cells and ECM. Flow is a critical determinant of vessel maintenance and durability. Lower panel: when insufficient angio- and arteriogenic factors are present and angiogenesis inhibitors are present, EC channels remain naked, leaky and fragile, are easily ruptured and bleed—conditions that reduce flow and result in vessel regression. A partial list of molecules is indicated; see text for additional information.

signals are present, Ang-2 causes EC death and vessel regression<sup>66,67</sup>. A precise balance of Tie-2 signals thus seems critical, as an activating Tie-2 mutation causes venous malformations that are composed of dilated, serpiginous endothelial channels covered by a variable amount of SMCs<sup>68</sup>.

Additional signaling molecules, such as members of the TGF- $\beta$  superfamily, contribute to the resolution and maturation phases of angiogenesis, but in a pleiotropic manner. TGF- $\beta$  family ligands stimulate type II receptors that, in turn, phosphorylate type I receptors (such as activin receptor-like kinase (ALK)) and activate the downstream signaling Smads<sup>69</sup>. Endoglin is a type III receptor, which facilitates binding of TGF- $\beta$ 1 to the type II receptors. Both pro- and antiangiogenic properties have been ascribed to TGF- $\beta$ 1, through effects on ECs and other cell types. At low doses, TGF- $\beta$ 1 contributes to the angiogenic switch by upregulating angiogenic factors and proteinases, whereas at high doses, TGF- $\beta$ 1 inhibits EC growth, promotes basement membrane reformation and stimulates SMC differentiation and recruitment. Hereditary hemorrhagic telangiectasia (HHT), characterized by telangiectasias and arterio-venous malformations, has been associated with loss-of-function mutations of endoglin (HHT-1) and ALK-1 (HHT-2)<sup>69</sup>. Because interpretations of the respective roles of ALK-1 (with Smad1 and Smad5) and ALK5 (with Smad2 and Smad3) in the activation or resolution phases of angiogenesis differ, the precise mechanisms of the vascular abnormalities of HHT lesions remain uncertain<sup>69-71</sup>. Nevertheless, an imbalance between vessel growth and maturation seems to cause the excessive fusion of capillary plexi into cavernous vessels and the hyperdilation of large vessels<sup>72</sup>. Mutations in the type II bone morphogenetic protein receptor (BMPR)-2 gene, also belonging to the TGF- $\beta$  superfamily, cause primary pulmonary hypertension, in which pulmonary arterioles become occluded by intravascular endothelial tumors<sup>73</sup>. By downregulating BMPR-1A (mediating BMPR-2 signaling), increased Ang-1 levels may further contribute to primary pulmonary hypertension by recruiting SMCs around pulmonary vessels<sup>74</sup>. In other primary pulmonary hypertension subjects, ECs acquire somatic mutations that lead to 'misguided angiogenesis'<sup>75,76</sup>.

### Collateral growth

Unlike distal capillaries, which distribute blood flow to individual cells, arteries provide bulk flow to the tissue and are therefore of utmost importance. When an artery is occluded, its vascular territory becomes ischemic. Because arterial systems are often interconnected by pre-existing collateral vessels, however, the collaterals can enlarge and salvage the ischemic region<sup>77</sup>. The mechanisms of angiogenesis and collateral growth differ significantly. Because of the large pressure differences between the perfusion territories, the increased shear stress activates ECs, which then recruit monocytes. These cells produce growth factors and

proteinases (uPA and MMPs), which enable SMCs to migrate and divide, explaining why depletion of monocytes impairs, whereas delivery of monocytes enhances, collateral growth<sup>78,79</sup>. Cytokines that attract monocytes or prolong their life span (such as monocyte chemoattractant protein (MCP)-1, granulocyte-macrophage colony-stimulating factor, TGF- $\beta$ 1 and tumor necrosis factor- $\alpha$ ) enhance collateral growth, whereas anti-inflammatory cytokines (such as IL-10) are inhibitory<sup>80-83</sup>. PlGF also enhances collateral growth, not only because it recruits monocytes, but also because it stimulates EC and SMC growth<sup>12,84</sup>. Delivery of acidic FGF, FGF-4 or basic FGF (together with PDGF-BB) stimulates collateral growth, in part by upregulating PDGFR expression<sup>85</sup>. VEGF alone seems to affect capillary angiogenesis more efficiently than collateral growth, explaining, at least in part, why results of clinical trials have not been more positive<sup>77,86</sup>. Coadministration of VEGF with additional molecules such as PDGF, PlGF or Ang-1 may enhance its therapeutic potential (ref. 58 and P.C., unpublished data). The identification of molecules

**Table 2 Diseases characterized or caused by insufficient angiogenesis or vessel regression**

Organ	Disease in mice or humans	Angiogenic mechanism
Nervous system	Alzheimer disease	Vasoconstriction, microvascular degeneration and cerebral angiopathy due to EC toxicity by amyloid- $\beta$ <sup>117</sup>
	Amyotrophic lateral sclerosis; diabetic neuropathy	Impaired perfusion and neuroprotection, causing motoneuron or axon degeneration due to insufficient VEGF production <sup>39</sup>
	Stroke	Correlation of survival with angiogenesis in brain <sup>118</sup> ; stroke due to arteriopathy (Notch-3 mutations) <sup>20</sup>
Blood vessels	Atherosclerosis	Characterized by impaired collateral vessel development <sup>119</sup>
	Hypertension	Microvessel rarefaction due to impaired vasodilation or angiogenesis <sup>105</sup>
	Diabetes	Characterized by impaired collateral growth <sup>120</sup> and angiogenesis in ischemic limbs <sup>121</sup> , but enhanced retinal neovascularization secondary to pericyte dropout
	Restenosis	Impaired re-endothelialization after arterial injury at old age <sup>122</sup>
Gastrointestinal	Gastric or oral ulcerations	Delayed healing due to production of angiogenesis inhibitors by pathogens <sup>123</sup> .
	Crohn disease	Characterized by mucosal ischemia
Skin	Hair loss	Retarded hair growth by angiogenesis inhibitors <sup>124</sup>
	Skin purpura, telangiectasia and venous lake formation	Age-dependent reduction of vessel number and maturation (SMC dropout) due to EC telomere shortening <sup>125</sup>
Reproductive system	Pre-eclampsia	EC dysfunction resulting in organ failure, thrombosis and hypertension due to deprivation of VEGF by soluble Flt-1 (ref. 126)
	Menorrhagia (uterine bleeding)	Fragility of SMC-poor vessels due to low Ang-1 production <sup>127</sup>
Lung	Neonatal respiratory distress	Insufficient lung maturation and surfactant production in premature mice due to reduced HIF-2 $\alpha$ and VEGF production <sup>128</sup>
	Pulmonary fibrosis, emphysema	Alveolar EC apoptosis upon VEGF inhibition <sup>129</sup>
Kidney	Nephropathy	Age-related vessel loss due to TSP-1 production <sup>130</sup>
Bone	Osteoporosis, impaired bone fracture healing	Impaired bone formation due to age dependent decline of VEGF-driven angiogenesis <sup>131</sup> ; angiogenesis inhibitors prevent fracture healing <sup>132</sup>

regulating collateral growth offers significant potential for the treatment of ischemic heart and limb disease.

### Leukocytes and angiogenesis

Inflammation- and immune-driven angiogenesis affect numerous disorders (Tables 1 and 2), in part because most leukocyte subtypes produce a myriad of angiogenic factors such as VEGF, PlGF, PDGF, basic FGF, Ang-2, epidermal growth factor, TGF- $\beta$ 1, MCP-1 and various interleukins and proteinases (trypsin, chymase, MMPs, heparanase and uPA; Fig. 1)<sup>87,88</sup>. Leukocytes affect many angiogenic processes. For instance, neutrophils and natural killer cells have been implicated in cyclical uterine angiogenesis, and in abnormal angiogenesis in endometriosis<sup>89</sup>, whereas tumor-associated macrophages promote cancer by releasing angiogenic factors and inducing tumor cells to release angiogenic factors<sup>90</sup>. Mast cells, when they encounter allergens and pathogens in the skin and mucosa, release vasoactive and angiogenic factors, thereby affecting autoimmune diseases in many organs. Mast cells also infiltrate skin carcinomas, where they hyperactivate angiogenesis through chymase-dependent activation of MMP-9 (ref. 91). Type I dendritic cells help eradicate tumors through immune stimulation and suppression of tumor angiogenesis<sup>92</sup>. Monocytes are a source of EPCs<sup>6</sup> and can differentiate into endothelial-like cells<sup>93</sup>. Because leukocytes also generate angiogenesis inhibitors, their overall role in initiating or terminating angiogenesis depends on the temporal and spatial balance of these modulators.

Leukocytes and vascular cells influence each other in other ways (Fig. 1). Angiogenic factors amplify the inflammatory process by recruiting leukocytes and affecting their function<sup>12</sup>. For instance, VEGF enhances, whereas TSP-1 and Ang-1 forestall, T-cell-dependent allograft arteriopathy by reducing leukocyte infiltration<sup>94</sup>. VEGF promotes cancer, not only by stimulating angiogenesis, but also by inhibiting the functional maturation of dendritic cells and enhancing adhesion of natural killer cells to tumor microvessels<sup>95,96</sup>. Other angiogenic molecules (such as PlGF, TGF- $\beta$ 1, PDGF and FGFs) also modulate leukocyte function<sup>12</sup>. Because of the significant involvement of leukocytes, anti-inflammatory drugs suppress pathological angiogenesis<sup>97</sup>. Another class of candidates are chemokines, which recruit leukocytes and directly stimulate ECs. These include growth-related oncogene, IL-8, stromal cell-derived factor-1, MCP-1 and others that bind CXCR2 and CXCR4 receptors<sup>98</sup>.

### Coagulation and angiogenesis

Fibrin-rich clot formation and platelet aggregation precede infiltration of blood vessels into a wound. Not surprisingly, therefore, hemostasis and angiogenesis are closely linked<sup>99–101</sup> (Fig. 1). Upon activation, platelets release large stores of angiogenic factors such as VEGF, PDGF, TGF- $\beta$ , IL-6, thrombin and sphingosine-1-phosphate. The latter stimulates the growth and stability of nascent vessels by tightening their junctions and recruiting mural cells<sup>102</sup>. Platelets also contain antiangiogenic factors (TSP-1, PF-4 and others) that may have a role in the resolution of angiogenesis once the wound has healed. The link between angiogenesis and hemostasis also has implications for cancer. Thromboembolism is a common cause of death in cancer patients. By covering tumor cells, platelets protect tumor emboli from immune surveillance and promote their lodging at distant metastatic sites. In many tumors, production of tissue factor, initiation of coagulation, and microvessel density are closely associated<sup>101</sup>. Tissue factor upregulates VEGF, downregulates TSP-1 and, by initiating coagulation, generates additional angiogenic pathways that are dependent on factor Xa, thrombin, the protease-activated receptors (PAR-1, PAR-2, PAR-3 and PAR-4) and fibrin<sup>100</sup>. The incidence of thrombosis in cancer patients treated with angiogenesis inhibitors may be attributable to EC dysfunction and death, platelet activation, the release of tumor

procoagulants and cytokines upon tumor lysis, and an inflammatory response<sup>99</sup>.

### Vessel regression

Vessel regression, a physiological mechanism to match perfusion with metabolic demand, occurs when the nascent vasculature consists of too many vessels. Vessel regression also constitutes the basis of many antiangiogenic therapeutic strategies. Abnormal vessel regression also contributes to the pathogenesis of numerous disorders, however. Several mechanisms shift the angiogenic switch from 'on' to 'off' (Fig. 2 and Table 2). Removal of angiogenic stimuli causes vessels to regress, as in tumors<sup>103</sup> and the heart<sup>104</sup>, especially when vessels have only been recently assembled and are still immature. When angiogenic stimuli are provided for a sufficient length of time, new vessels mature and persist for months, even after the angiogenic stimulus is withdrawn<sup>104</sup>. Flow may have an important role in determining whether neovessels regress or persist. By affecting several factors (including MMPs, PDGF, basic FGF, integrins and nitric oxide), flow stimulates hyperplasia of ECs and SMCs, and induces the reorganization of endothelial junctions and the deposition of ECM—all of which contribute to vessel maturation. Thus, insufficient perfusion may lead to regression, whereas sufficient perfusion promotes vessel persistence. An abnormal sensitivity of small arterioles to vasoconstrictor stimuli may lead to functional constriction and subsequent structural rarefaction of nonperfused 'ghost arterioles' in hypertension<sup>105</sup>. Pericytes also determine the susceptibility of vessels to regression. Indeed, once vessels are surrounded by pericytes, they become resistant to oxygen-induced regression<sup>103</sup>. Delivery of PlGF or VEGF with PDGF-BB causes vessel maturation and results in the persistence of stable, durable vessels for more than a year<sup>12,58</sup>. In contrast, disruption of endothelial-pericyte associations results in the regression of vessels<sup>106</sup>.

Angiogenesis inhibitors also contribute to vessel regression. TSP-1 inhibits angiogenesis through direct effects on ECs and indirect effects on growth factor mobilization or activation<sup>107</sup>. Upregulation of endogenous TSP-1 and TSP-2 contributes to the resolution of angiogenesis and vessel stabilization after ischemia, and forced overexpression of TSP-1 or TSP-2 in cancer cells results in reduced tumor vascularization and tumor growth<sup>107</sup>. There are more angiogenesis inhibitors, however. When VEGF levels are low, Ang-2 marks regressing vessels<sup>108</sup>; interferons exert angiostatic effects by lowering the expression of basic FGF and VEGF. Macrophages (such as hyalocytes in the eye) contribute to vessel regression by releasing TGF- $\beta$ 1 (ref. 109). Inhibitory PAS domain protein, a splice variant of HIF-3 $\alpha$ , functions as a dominant-negative regulator of hypoxia-induced angiogenesis to maintain an avascular phenotype in certain tissues<sup>110</sup>. Additional inhibitors include chemokines binding CXCR3 (such as PF-4, Mig, interferon-inducible protein-10 and others)<sup>98</sup>, soluble receptors (Flt-1 and Tie-2), clotting antagonists and others. A growing list of inhibitors is being discovered, including cleavage products of matrix components (such as arresten, constatin and tumstatin from collagen IV; vastatin from collagen VIII; restin from collagen XV; and endostatin from collagen XVIII), proteinases or enzymes (such as PEX from MMP2; mini-TrpRS from tryptophanyl-tRNA synthetase) or plasma proteins (such as angiostatin from plasminogen; 16K prolactin from prolactin; and fragments of several serpins)<sup>111</sup>. The endogenous roles of many of these cleavage products in physiological and pathological angiogenesis remain enigmatic. Nevertheless, they offer opportunities to suppress tumor angiogenesis and growth when administered.

### Conclusion

Historically, angiogenesis was initially only implicated in cancer, arthritis and psoriasis. In recent years it has, however, become increasingly evident that excessive, insufficient or abnormal angiogenesis contributes to the

pathogenesis of many more disorders. Ongoing clinical trials reveal that both pro- and antiangiogenic treatments with single angiogenic molecules is more challenging than anticipated, and monotherapy with a single angiogenesis inhibitor may not suffice to combat the myriad of angiogenic factors produced by cancer cells. This may not be surprising, however, when one considers that building new, functional and durable vessels requires a complex interplay of multiple molecular signals. The challenge for the coming years is thus to define the molecular basis and pathways of angiogenic disorders in greater detail and in a more integrated manner, so that the excitement of the science can be converted into the development of efficient, safe therapies.

#### ACKNOWLEDGMENTS

The author thanks all members of the Center for Transgene Technology and Gene Therapy and all external collaborators, and A. Vandenhoek for artwork.

- Luttun, A., Carmeliet, G. & Carmeliet, P. Vascular progenitors: from biology to treatment. *Trends Cardiovasc. Med.* **12**, 88–96 (2002).
- Rafii, S., Lyden, D., Benezra, R., Hattori, K. & Heissig, B. Vascular and haematopoietic stem cells: novel targets for anti-angiogenesis therapy? *Nat. Rev. Cancer* **2**, 826–835 (2002).
- Asahara, T. & Isner, J.M. Endothelial progenitor cells for vascular regeneration. *J. Hematother. Stem Cell Res.* **11**, 171–178 (2002).
- Mikkola, H.K. & Orkin, S.H. The search for the hemangioblast. *J. Hematother. Stem Cell Res.* **11**, 9–17 (2002).
- Reyes, M. *et al.* Origin of endothelial progenitors in human postnatal bone marrow. *J. Clin. Invest.* **109**, 337–346 (2002).
- Rehman, J., Li, J., Orschemel, C.M. & March, K.L. Peripheral blood “endothelial progenitor cells” are derived from monocyte/macrophages and secrete angiogenic growth factors. *Circulation* **107**, 1164–1169 (2003).
- Takakura, N. *et al.* A role for hematopoietic stem cells in promoting angiogenesis. *Cell* **102**, 199–209 (2000).
- Grant, M.B. *et al.* Adult hematopoietic stem cells provide functional hemangioblast activity during retinal neovascularization. *Nat. Med.* **8**, 607–612 (2002).
- Gerber, H.P. *et al.* VEGF regulates haematopoietic stem cell survival by an internal autocrine loop mechanism. *Nature* **417**, 954–958 (2002).
- Hattori, K. *et al.* Placental growth factor reconstitutes hematopoiesis by recruiting VEGFR1<sup>+</sup> stem cells from bone-marrow microenvironment. *Nat. Med.* **8**, 841–849 (2002).
- Lyden, D. *et al.* Impaired recruitment of bone-marrow-derived endothelial and hematopoietic precursor cells blocks tumor angiogenesis and growth. *Nat. Med.* **7**, 1194–1201 (2001).
- Luttun, A. *et al.* Revascularization of ischemic tissues by PIGF treatment, and inhibition of tumor angiogenesis, arthritis and atherosclerosis by anti-Flt1. *Nat. Med.* **8**, 831–840 (2002).
- Rafii, S. & Lyden, D. Therapeutic stem and progenitor cell transplantation for organ vascularization and regeneration. *Nat. Med.* **9**, 702–712 (2003).
- Carmeliet, P. Developmental biology. One cell, two fates. *Nature* **408**, 43–45 (2000).
- Lawson, N.D. *et al.* Notch signaling is required for arterial-venous differentiation during embryonic vascular development. *Development* **128**, 3675–3683 (2001).
- Zhong, T.P., Childs, S., Leu, J.P. & Fishman, M.C. Gridlock signalling pathway fashions the first embryonic artery. *Nature* **414**, 216–220 (2001).
- Lawson, N.D., Vogel, A.M. & Weinstein, B.M. Sonic hedgehog and vascular endothelial growth factor act upstream of the Notch pathway during arterial endothelial differentiation. *Dev. Cell* **3**, 127–136 (2002).
- Stalmans, I. *et al.* Arteriolar and venular patterning in retinas of mice selectively expressing VEGF isoforms. *J. Clin. Invest.* **109**, 327–336 (2002).
- Visconti, R.P., Richardson, C.D. & Sato, T.N. Orchestration of angiogenesis and arteriovenous contribution by angiopoietins and vascular endothelial growth factor (VEGF). *Proc. Natl. Acad. Sci. USA* **99**, 8219–8224 (2002).
- Kalimo, H., Ruchoux, M.M., Viitanen, M. & Kalaria, R.N. CADASIL: a common form of hereditary arteriopathy causing brain infarcts and dementia. *Brain Pathol.* **12**, 371–384 (2002).
- Mukoyama, Y.S., Shin, D., Britsch, S., Taniguchi, M. & Anderson, D.J. Sensory nerves determine the pattern of arterial differentiation and blood vessel branching in the skin. *Cell* **109**, 693–705 (2002).
- Cleaver, O. & Melton, D.A. Endothelial signaling during development. *Nat. Med.* **9**, 1–4 (2003).
- Jain, R.K. & Munn, L.L. Leaky vessels? Call Ang1! *Nat. Med.* **6**, 131–132 (2000).
- Suri, C. *et al.* Increased vascularization in mice overexpressing angiopoietin-1. *Science* **282**, 468–471 (1998).
- LeCouter, J. *et al.* Identification of an angiogenic mitogen selective for endocrine gland endothelium. *Nature* **412**, 877–884 (2001).
- Ruoslahti, E. Drug targeting to specific vascular sites. *Drug Discov. Today* **7**, 1138–1143 (2002).
- Sood, A.K., Fletcher, M.S. & Hendrix, M.J. The embryonic-like properties of aggressive human tumor cells. *J. Soc. Gynecol. Investig.* **9**, 2–9 (2002).
- Wang, H.U., Chen, Z.F. & Anderson, D.J. Molecular distinction and angiogenic interaction between embryonic arteries and veins revealed by ephrin-B2 and its receptor EphB4. *Cell* **93**, 741–753 (1998).
- Gerety, S.S., Wang, H.U., Chen, Z.F. & Anderson, D.J. Symmetrical mutant phenotypes of the receptor EphB4 and its specific transmembrane ligand ephrin-B2 in cardiovascular development. *Mol. Cell* **4**, 403–414 (1999).
- Zhang, X.Q. *et al.* Stromal cells expressing ephrin-B2 promote the growth and sprouting of ephrin-B2(+) endothelial cells. *Blood* **98**, 1028–1037 (2001).
- Gale, N.W. *et al.* Ephrin-B2 selectively marks arterial vessels and neovascularization sites in the adult, with expression in both endothelial and smooth-muscle cells. *Dev. Biol.* **230**, 151–160 (2001).
- Shin, D. *et al.* Expression of ephrinB2 identifies a stable genetic difference between arterial and venous vascular smooth muscle as well as endothelial cells, and marks subsets of microvessels at sites of adult neovascularization. *Dev. Biol.* **230**, 139–150 (2001).
- Stalmans, I. *et al.* VEGF: A modifier of the del22q11 (DiGeorge) syndrome? *Nat. Med.* **9**, 173–182 (2003).
- Loughna, S. & Sato, T.N. A combinatorial role of angiopoietin-1 and orphan receptor TIE1 pathways in establishing vascular polarity during angiogenesis. *Mol. Cell* **7**, 233–239 (2001).
- Pugh, C.W. & Ratcliffe, P.J. Regulation of angiogenesis by hypoxia: role of the HIF system. *Nat. Med.* **9**, 677–684 (2003).
- Ferrara, N., Gerber, H.-P., LeCouter, J. & Lin, R. The biology of VEGF and its receptors. *Nat. Med.* **9**, 669–676 (2003).
- Carmeliet, P. *et al.* Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele. *Nature* **380**, 435–439 (1996).
- Ferrara, N. *et al.* Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene. *Nature* **380**, 439–442 (1996).
- Oosthuysen, B. *et al.* Deletion of the hypoxia-response element in the vascular endothelial growth factor promoter causes motor neuron degeneration. *Nat. Genet.* **28**, 131–138 (2001).
- Carmeliet, P. *et al.* Synergism between vascular endothelial growth factor and placental growth factor contributes to angiogenesis and plasma extravasation in pathological conditions. *Nat. Med.* **7**, 575–583 (2001).
- Carmeliet, P. *et al.* Targeted deficiency or cytosolic truncation of the VE-cadherin gene in mice impairs VEGF-mediated endothelial survival and angiogenesis. *Cell* **98**, 147–157 (1999).
- Corada, M. *et al.* A monoclonal antibody to vascular endothelial-cadherin inhibits tumor angiogenesis without side effects on endothelial permeability. *Blood* **100**, 905–911 (2002).
- Thurston, G. *et al.* Angiopoietin-1 protects the adult vasculature against plasma leakage. *Nat. Med.* **6**, 460–463 (2000).
- Simon, A.M. & McWhorter, A.R. Vascular abnormalities in mice lacking the endothelial gap junction proteins connexin37 and connexin40. *Dev. Biol.* **251**, 206–220 (2002).
- Hangai, M. *et al.* Matrix metalloproteinase-9-dependent exposure of a cryptic migratory control site in collagen is required before retinal angiogenesis. *Am. J. Pathol.* **161**, 1429–1437 (2002).
- Hynes, R.O. A reevaluation of integrins as regulators of angiogenesis. *Nat. Med.* **8**, 918–921 (2002).
- Hood, J.D. & Cheresch, D.A. Role of integrins in cell invasion and migration. *Nat. Rev. Cancer* **2**, 91–100 (2002).
- Pepper, M.S. Extracellular proteolysis and angiogenesis. *Thromb. Haemost.* **86**, 346–355 (2001).
- Jackson, C. Matrix metalloproteinases and angiogenesis. *Curr. Opin. Nephrol. Hypertens.* **11**, 295–299 (2002).
- Luttun, A., Dewerchin, M., Collen, D. & Carmeliet, P. The role of proteinases in angiogenesis, heart development, restenosis, atherosclerosis, myocardial ischemia, and stroke: insights from genetic studies. *Curr. Atheroscler. Rep.* **2**, 407–416 (2000).
- Qi, J.H. *et al.* A novel function for tissue inhibitor of metalloproteinases-3 (TIMP3): inhibition of angiogenesis by blockage of VEGF binding to VEGF receptor-2. *Nat. Med.* **9**, 407–415 (2003).
- Blasi, F. & Carmeliet, P. uPAR: a versatile signalling orchestrator. *Nat. Rev. Mol. Cell Biol.* **3**, 932–943 (2002).
- Bajou, K. *et al.* Absence of host plasminogen activator inhibitor 1 prevents cancer invasion and vascularization. *Nat. Med.* **4**, 923–928 (1998).
- Jain, R.K. Molecular regulation of vessel maturation. *Nat. Med.* **9**, 685–693 (2003).
- Hellstrom, M. *et al.* Lack of pericytes leads to endothelial hyperplasia and abnormal vascular morphology. *J. Cell Biol.* **153**, 543–553 (2001).
- Abramsson, A. *et al.* Analysis of mural cell recruitment to tumor vessels. *Circulation* **105**, 112–117 (2002).
- Dinehart, S.M., Kincannon, J. & Geronemus, R. Hemangiomas: evaluation and treatment. *Dermatol. Surg.* **27**, 475–485 (2001).
- Richardson, T.P., Peters, M.C., Ennett, A.B. & Mooney, D.J. Polymeric system for dual growth factor delivery. *Nat. Biotechnol.* **19**, 1029–1034 (2001).
- Cao, R. *et al.* Angiogenesis stimulated by PDGF-CC, a novel member in the PDGF family, involves activation of PDGFR- $\alpha$  and - $\beta$  receptors. *FASEB J.* **16**, 1575–1583 (2002).
- Takagi, H. *et al.* Potential role of the angiopoietin/tie2 system in ischemia-induced retinal neovascularization. *Invest. Ophthalmol. Mol. Vis. Sci.* **44**, 393–402 (2003).
- Shim, W.S. *et al.* Angiopoietin 1 promotes tumor angiogenesis and tumor vessel plasticity of human cervical cancer in mice. *Exp. Cell Res.* **279**, 299–309 (2002).
- Hattori, K. *et al.* Vascular endothelial growth factor and angiopoietin-1 stimulate postnatal hematopoiesis by recruitment of vasculogenic and hematopoietic stem cells. *J. Exp. Med.* **193**, 1005–1014 (2001).
- Ahmad, S.A. *et al.* The effects of angiopoietin-1 and -2 on tumor growth and angiogenesis in human colon cancer. *Cancer Res.* **61**, 1255–1259 (2001).

64. Carlson, T.R., Feng, Y., Maisonnier, P.C., Mrksich, M. & Morla, A.O. Direct cell adhesion to the angiotensins mediated by integrins. *J. Biol. Chem.* **276**, 26516–26525 (2001).
65. Gale, N.W. *et al.* Angiotensin-2 is required for postnatal angiogenesis and lymphatic patterning, and only the latter role is rescued by Angiotensin-1. *Dev. Cell* **3**, 411–23 (2002).
66. Maisonnier, P.C. *et al.* Angiotensin-2, a natural antagonist for Tie2 that disrupts *in vivo* angiogenesis. *Science* **277**, 55–60 (1997).
67. Hackett, S.F., Wiegand, S., Yancopoulos, G. & Campochiaro, P.A. Angiotensin-2 plays an important role in retinal angiogenesis. *J. Cell. Physiol.* **192**, 182–187 (2002).
68. Vikkula, M. *et al.* Vascular dysmorphogenesis caused by an activating mutation in the receptor tyrosine kinase TIE2. *Cell* **87**, 1181–1190 (1996).
69. van den Driesche, S., Mummery, C.L. & Westermann, C.J. Hereditary hemorrhagic telangiectasia: an update on transforming growth factor  $\beta$  signaling in vasculogenesis and angiogenesis. *Cardiovasc. Res.* **58**, 20–31 (2003).
70. Lamouille, S., Mallet, C., Feige, J.J. & Bailly, S. Activin receptor-like kinase 1 is implicated in the maturation phase of angiogenesis. *Blood* **100**, 4495–4501 (2002).
71. Goumans, M.J. *et al.* Balancing the activation state of the endothelium via two distinct TGF- $\beta$  type I receptors. *EMBO J.* **21**, 1743–1753 (2002).
72. Srinivasan, S. *et al.* A mouse model for hereditary hemorrhagic telangiectasia (HHT) type 2. *Hum. Mol. Genet.* **12**, 473–482 (2003).
73. Humbert, M. & Trembath, R.C. Genetics of pulmonary hypertension: from bench to bedside. *Eur. Respir. J.* **20**, 741–749 (2002).
74. Du, L. *et al.* Signaling molecules in nonfamilial pulmonary hypertension. *N. Engl. J. Med.* **348**, 500–509 (2003).
75. Voelkel, N.F. *et al.* Janus face of vascular endothelial growth factor: the obligatory survival factor for lung vascular endothelium controls precapillary artery remodeling in severe pulmonary hypertension. *Crit. Care Med.* **30**, S251–S256 (2002).
76. Yeager, M.E., Halley, G.R., Golpon, H.A., Voelkel, N.F. & Tuder, R.M. Microsatellite instability of endothelial cell growth and apoptosis genes within plexiform lesions in primary pulmonary hypertension. *Circ. Res.* **88**, E2–E11 (2001).
77. Helisch, A. & Schaper, W. Arteriogenesis: the development and growth of collateral arteries. *Microcirculation* **10**, 83–97 (2003).
78. Kamihata, H. *et al.* Improvement of collateral perfusion and regional function by implantation of peripheral blood mononuclear cells into ischemic hibernating myocardium. *Arterioscler. Thromb. Biol.* **22**, 1804–1810 (2002).
79. Heil, M. *et al.* Blood monocyte concentration is critical for enhancement of collateral artery growth. *Am. J. Physiol. Heart Circ. Physiol.* **283**, H2411–H2419 (2002).
80. van Royen, N. *et al.* Exogenous application of transforming growth factor  $\beta$  stimulates arteriogenesis in the peripheral circulation. *FASEB J.* **16**, 432–434 (2002).
81. Buschmann, I.R. *et al.* GM-CSF: a strong arteriogenic factor acting by amplification of monocyte function. *Atherosclerosis* **159**, 343–356 (2001).
82. Voskuil, M. *et al.* Modulation of collateral artery growth in a porcine hindlimb ligation model using MCP-1. *Am. J. Physiol. Heart Circ. Physiol.* **284**, H1422–H1428 (2003).
83. Hoefler, I.E. *et al.* Direct evidence for tumor necrosis factor- $\alpha$  signaling in arteriogenesis. *Circulation* **105**, 1639–1641 (2002).
84. Pipp, F. *et al.* VEGFR-1-selective VEGF homologue PIGF is arteriogenic: evidence for a monocyte-mediated mechanism. *Circ. Res.* **92**, 378–385 (2003).
85. Cao, R. *et al.* Angiogenic synergism, vascular stability and improvement of hind-limb ischemia by a combination of PDGF-BB and FGF-2. *Nat. Med.* (2003).
86. Isner, J.M. Myocardial gene therapy. *Nature* **415**, 234–239 (2002).
87. Vacca, A. *et al.* Human lymphoblastoid cells produce extracellular matrix-degrading enzymes and induce endothelial cell proliferation, migration, morphogenesis, and angiogenesis. *Int. J. Clin. Lab. Res.* **28**, 55–68 (1998).
88. Norrby, K. Mast cells and angiogenesis. *APMIS* **110**, 355–371 (2002).
89. Li, X.F. *et al.* Angiogenic growth factor messenger ribonucleic acids in uterine natural killer cells. *J. Clin. Endocrinol. Metab.* **86**, 1823–1834 (2001).
90. Sica, A., Saccani, A. & Mantovani, A. Tumor-associated macrophages: a molecular perspective. *Int. Immunopharmacol.* **2**, 1045–1054 (2002).
91. Coussens, L.M. *et al.* Inflammatory mast cells up-regulate angiogenesis during squamous epithelial carcinogenesis. *Genes Dev.* **13**, 1382–1397 (1999).
92. Banachereau, J. & Steinman, R.M. Dendritic cells and the control of immunity. *Nature* **392**, 245–252 (1998).
93. Schmeisser, A. & Strasser, R.H. Phenotypic overlap between hematopoietic cells with suggested angioblastic potential and vascular endothelial cells. *J. Hematother. Stem Cell Res.* **11**, 69–79 (2002).
94. Nykanen, A.I. *et al.* Angiotensin-1 protects against the development of cardiac allograft arteriosclerosis. *Circulation* **107**, 1308–1314 (2003).
95. Melder, R.J. *et al.* During angiogenesis, vascular endothelial growth factor and basic fibroblast growth factor regulate natural killer cell adhesion to tumor endothelium. *Nat. Med.* **2**, 992–997 (1996).
96. Carbone, J.E. & Ohm, D.P. Immune dysfunction in cancer patients. *Oncology (Huntington)* **16**, 11–18 (2002).
97. Dermond, O. & Ruegg, C. Inhibition of tumor angiogenesis by non-steroidal anti-inflammatory drugs: emerging mechanisms and therapeutic perspectives. *Drug Resist. Update* **4**, 314–321 (2001).
98. Bernardini, G. *et al.* Analysis of the role of chemokines in angiogenesis. *J. Immunol. Meth.* **273**, 83–101 (2003).
99. Trikha, M. & Nakada, M.T. Platelets and cancer: implications for antiangiogenic therapy. *Semin. Thromb. Hemost.* **28**, 39–44 (2002).
100. Carmeliet, P. Biomedicine. Clotting factors build blood vessels. *Science* **293**, 1602–1604 (2001).
101. Fernandez, P.M. & Rickles, F.R. Tissue factor and angiogenesis in cancer. *Curr. Opin. Hematol.* **9**, 401–406 (2002).
102. English, D., Brindley, D.N., Spiegel, S. & Garcia, J.G. Lipid mediators of angiogenesis and the signalling pathways they initiate. *Biochim. Biophys. Acta* **1582**, 228–239 (2002).
103. Benjamin, L.E., Golijanin, D., Itin, A., Podes, D. & Keshet, E. Selective ablation of immature blood vessels in established human tumors follows vascular endothelial growth factor withdrawal. *J. Clin. Invest.* **103**, 159–165 (1999).
104. Dor, Y. *et al.* Conditional switching of VEGF provides new insights into adult neovascularization and pro-angiogenic therapy. *EMBO J.* **21**, 1939–1947 (2002).
105. Boudier, H.A. Arteriole and capillary remodelling in hypertension. *Drugs* **58** (suppl. 1), 37–40 (1999).
106. Benjamin, L.E., Hemo, I. & Keshet, E. A plasticity window for blood vessel remodelling is defined by pericyte coverage of the preformed endothelial network and is regulated by PDGF-B and VEGF. *Development* **125**, 1591–1598 (1998).
107. Vailhe, B. & Feige, J.J. Thrombospondins as anti-angiogenic therapeutic agents. *Curr. Pharm. Des.* **9**, 583–588 (2003).
108. Holash, J. *et al.* Vessel cooption, regression, and growth in tumors mediated by angiotensins and VEGF. *Science* **284**, 1994–1998 (1999).
109. Schonfeld, C.L. Hyalocytes inhibit retinal pigment epithelium cell proliferation *in vitro*. *Ger. J. Ophthalmol.* **5**, 224–228 (1996).
110. Makino, Y., Kanopka, A., Wilson, W.J., Tanaka, H. & Poellinger, L. Inhibitory PAS domain protein (IPAS) is a hypoxia-inducible splicing variant of the hypoxia-inducible factor-3 $\alpha$  locus. *J. Biol. Chem.* **277**, 32405–32408 (2002).
111. D'Amore, P.A. & Ng, Y.S. Tales of the cryptic: unveiling more angiogenesis inhibitors. *Trends Mol. Med.* **8**, 313–315 (2002).
112. Meyer, M. *et al.* A novel vascular endothelial growth factor encoded by Orf virus, VEGF-E, mediates angiogenesis via signalling through VEGFR-2 (KDR) but not VEGFR-1 (Flt-1) receptor tyrosine kinases. *EMBO J.* **18**, 363–374 (1999).
113. Harada, K., Lu, S., Chisholm, D.M., Syrjanen, S. & Schor, A.M. Angiogenesis and vasodilation in skin warts. Association with HPV infection. *Anticancer Res.* **20**, 4519–4523 (2000).
114. Barillari, G. & Enslin, B. Angiogenic effects of extracellular human immunodeficiency virus type 1 Tat protein and its role in the pathogenesis of AIDS-associated Kaposi's sarcoma. *Clin. Microbiol. Rev.* **15**, 310–326 (2002).
115. Ruppnick, M.A. *et al.* Adipose tissue mass can be regulated through the vasculature. *Proc. Natl. Acad. Sci. USA* **99**, 10730–10735 (2002).
116. Hackett, S.F. *et al.* Angiotensin 2 expression in the retina: upregulation during physiologic and pathologic neovascularization. *J. Cell Physiol.* **184**, 275–284 (2000).
117. De La Torre, J.C. Alzheimer's disease: How does it start? *J. Alzheimers Dis.* **4**, 497–512 (2002).
118. Krupinski, J., Kaluza, J., Kumar, P., Kumar, S. & Wang, J.M. Role of angiogenesis in patients with cerebral ischemic stroke. *Stroke* **25**, 1794–1798 (1994).
119. Van Belle, E. *et al.* Hypercholesterolemia attenuates angiogenesis but does not preclude augmentation by angiogenic cytokines. *Circulation* **96**, 2667–2674 (1997).
120. Waltenberger, J. Impaired collateral vessel development in diabetes: potential cellular mechanisms and therapeutic implications. *Cardiovasc. Res.* **49**, 554–560 (2001).
121. Rivard, A. *et al.* Rescue of diabetes-related impairment of angiogenesis by intramuscular gene therapy with adeno-VEGF. *Am. J. Pathol.* **154**, 355–363 (1999).
122. Gennaro, G., Menard, C., Michaud, S.E. & Rivard, A. Age-dependent impairment of reendothelialization after arterial injury: role of vascular endothelial growth factor. *Circulation* **107**, 230–233 (2003).
123. Jenkinson, L., Bardhan, K.D., Atherton, J. & Kalia, N. *Helicobacter pylori* prevents proliferative stage of angiogenesis *in vitro*: role of cytokines. *Dig. Dis. Sci.* **47**, 1857–1862 (2002).
124. Yano, K., Brown, L.F. & Detmar, M. Control of hair growth and follicle size by VEGF-mediated angiogenesis. *J. Clin. Invest.* **107**, 409–417 (2001).
125. Chang, E., Yang, J., Nagavarapu, U. & Herron, G.S. Aging and survival of cutaneous microvasculature. *J. Invest. Dermatol.* **118**, 752–758 (2002).
126. Maynard, S.E. *et al.* Excess placental soluble fms-like tyrosine kinase 1 (sFlt1) may contribute to endothelial dysfunction, hypertension, and proteinuria in preeclampsia. *J. Clin. Invest.* **111**, 649–658 (2003).
127. Hewett, P. *et al.* Down-regulation of angiotensin-1 expression in menorrhagia. *Am. J. Pathol.* **160**, 773–780 (2002).
128. Compernelle, V. *et al.* Loss of HIF-2 $\alpha$  and inhibition of VEGF impair fetal lung maturation, whereas treatment with VEGF prevents fatal respiratory distress in premature mice. *Nat. Med.* **8**, 702–710 (2002).
129. Kasahara, Y. *et al.* Inhibition of VEGF receptors causes lung cell apoptosis and emphysema. *J. Clin. Invest.* **106**, 1311–1319 (2000).
130. Kang, D.H. *et al.* Impaired angiogenesis in the aging kidney: vascular endothelial growth factor and thrombospondin-1 in renal disease. *Am. J. Kidney Dis.* **37**, 601–611 (2001).
131. Martinez, P., Esbrit, P., Rodrigo, A., Alvarez-Arroyo, M.V. & Martinez, M.E. Age-related changes in parathyroid hormone-related protein and vascular endothelial growth factor in human osteoblastic cells. *Osteoporos. Int.* **13**, 874–881 (2002).
132. Yin, G. *et al.* Endostatin gene transfer inhibits joint angiogenesis and pannus formation in inflammatory arthritis. *Mol. Ther.* **5**, 547–554 (2002).